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## =Abstract=

### The usefulness of the genetic markers at the low-density lipoprotein receptor gene locus for the genetic diagnosis of familial hypercholesterolemia

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**Background :** Familial hypercholesterolemia(FH) is an autosomal dominant metabolic disorder caused by the mutation in low density lipoprotein receptor(LDLR) gene. However, direct genetic diagnosis of LDLR gene mutation is not easily available because more than 300 mutations have been described in LDLR gene of FH patients. Therefore indirect genetic diagnosis using the genetic markers can be used to follow the inheritance of defective gene in FH families. The purpose of this study was to evaluate the usefulness of indirect genetic markers for detecting identical-by-descent LDLR gene abnormalities in FH families.

**Methods :** We examined the allele frequency, heterozygosity, polymorphism information content(PIC) of each genetic markers(D19S394, *Taq* I, *Hinc* II, *Ava* II, ATn, D19S221) in 94 unrelated healthy subjects. The genetic polymorphic haplotypes in 3 FH families were also determined.

**Results :** The heterozygosity and PIC values of RFLP's(*Taq* I, *Hinc* II, *Ava* II) were 0.51/0.344, 0.25/0.223, 0.28/0.233 and microsatellite markers(D19S394, ATn, D19S221) were 0.64/0.558, 0.56/0.455, 0.60/0.475. *Hinc* II and *Ava* II were significantly linked( $|D|=0.72$ ,  $p<0.05$ ). The cumulative PIC values of *Taq* I+*Hinc* II, *Taq* I+*Hinc* II+ATn, D19S394+ATn were 0.520, 0.814, 0.813, respectively.

When applied in the FH pedigree, the genetic diagnosis using only one marker was not available in most cases. However, combination of two or more genetic markers could successfully discriminate

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the affected and unaffected members in FH families. Among the several combinations of the genetic markers, the combination of D19S394 and ATn was supposed to be the most effective and informative. Because one case of recombination was suspected in D19S221 allele, it was thought to be carefully used for genetic diagnosis of FH.

**Conclusion** : We concluded that indirect genetic diagnosis using intragenic or extragenic genetic markers was useful for detecting identical-by-descent LDLR gene abnormalities in FH families and the most effective and informative combination of genetic marker seemed to be D19S394 and ATn. (Korean J Med 58:283- 292, 2000)

**Key Words** : Familial hypercholesterolemia, Genetic marker

FH: <http://www.ucl.ac.uk/fh>. LDLR

가 (familial hypercholesterolemia, FH) 25 FH 18가 LDLR 가

(low-density lipoprotein receptor, LDLR) . LDLR 가

1-3, FH 500- 가

1000 1 ,

2-4 가 LDL 20-30

FH , (genetic marker)

가 LDL , (triglyceride)

2a (hyperlipoproteinemia) ,

tendon xanthoma, 가 (identical-by-descent)

FH 가

2a FH FH

(polygenic hypercholesterolemia) 가

75% tendon xanthoma , (restriction

10% 가 가 fragment length polymorphism, RFLP)

FH 가 FH heterozygosity, polymorphism information content (PIC)

LDLR RFLP가

LDLR 19 short arm 89 tandem repeat sequence

18 exon , 1984 Yamamoto microsatellite marker 가

cloning LDLR FH

가 FH 46, 10 11

300 가 LDLR 가 LDLR *Taq* I, *Hinc* II, *Ava* II RFLP

( Database of LDLR gene mutations in exon18 3' (AT)n tandem repeat

**Table 1. Primer sequences for PCR of microsatellite markers**

Marker	Oligonucleotide sequence	Tandem repeat
D19S394	5'-CCA AGA TCC TCT CCT GAG GT-3' 5'-CTT TCT ACC TTT CTA TCA TCT GTC A-3'	GATA
ATn	5'-CAC TTT GTA TAT TGG TTG AAA CTG T-3' 5'-CAC TGA ACA AAT ACA GCA ACC AGG G-3'	AT
D19S221	5'-GCA AGA CTC TGA CTC AAC AAA A-3' 5'-CAT AGA GAT CAA TGG CAT GAA A-3'	CA

**Table 2. Primer sequences for PCR and the fragment sizes of RFLP markers**

Marker	Oligonucleotide sequence	Fragment size
<i>Taq</i> I (intron1)	5'-ATC TGA CGA GGA AAA CTG-3' 5'-GGC CAC AGC TGG AAA ACA-3'	600 and 450 bp
<i>Hinc</i> II (exon12)	5'-TCT GGG ACT GGC ATC AGC AC-3' 5'-AGA TAG GCC GGT GGA TTC AC-3'	133, 99, 57 and 53 bp
<i>Ava</i> II (exon13)	5'-NNN TTG CTG CCT GTT TAG-3' 5'-GTT CCT CCA AAG TTC CAA CC-3'	128 and 75 bp

N: A, C, G or T. The nucleotides were randomly applied because the 5' end of the sense primer was not informative.

(ATn) LDLR (intragenic) FH Weiss  
D19S394 (GATA tetranucleotide repeat), D19S221 가 (proband)  
(CA dinucleotide repeat) (extragenic) 가  
가 LDL- 160 mg/dl  
가 , 가 , tendon xanthoma  
12, 13. (first degree relative)가 60  
가 FH FH  
가 .  
2. DNA  
FH 가  
1. 7 ml EDTA가 가  
가 , genomic DNA Wizard DNA purification system<sup>®</sup>  
가 94 (Promega Co., Madison, Wisconsin, U.S.A.)  
allele , heterozy-  
gosity, PIC FH 가 15  
3. Polymerase chain reaction(PCR) PCR  
FH 가  
D19S394, *Taq* I, *Hinc* II, *Ava* II, ATn,  
(lipid lowering agent) D19S221 . D19S394 LDLR 5'  
250 kb telomere *Taq* I, *Hinc*

**Table 3. PCR conditions used to amplify the sequences of genetic markers**

Marker	PCR condition				
	Annealing		Extension		MgCl <sub>2</sub>
D19S394	55	1min	72	1min	1.5mM
<i>Taq</i> I	60	30sec	68	2min	1.0mM
<i>Hinc</i> II	65	1min	72	1min	1.5mM
<i>Ava</i> II	65	1min	72	1min	1.5mM
ATn	55	1min	72	1min	1.5mM
D19S221	63	1min	72	1min	1.5mM

II, *Ava* II RFLP intron 1, exon 12, Inc., Branchburg, New Jersey, U.S.A.)  
 exon 13 . ATn exon 18 3' . 95 10 *Taq* polymerase  
 D19S221 LDLR 3' 1.3 94 1 DNA (denaturation)  
 Mb centromere . PCR annealing extension  
 genomic DNA (template) 35 72 5 final extension  
 (RFLP microsatellite marker) .  
 primer PCR (Table 7) microsatellite marker sense  
 1, Table 2). primer T4 polynucleotide kinase [ - 3<sup>2</sup>P]ATP end labelling PCR  
 PCR 20 µl DNA 50 ng, sense , PCR 94 5 denaturing  
 antisense primer 10 pmole , 62.5 µM of dNTPs, 6% polyacrylamide gel .  
 45 mM Tris-HCl (pH 8.8), 11 mM ammonium sulfate, phosphor- imaging plate reader (Bio- imaging  
 6.7 mM -mercaptoethanol, 4.5 µM EDTA, PCR analyser system 2500<sup>®</sup>, Fuji Film Co., Tokyo, Japan)  
 MgCl<sub>2</sub> (Table 3), 0.4 U *Taq* PCR  
 polymerase (AmpliTaq Gold<sup>®</sup>, Roche Molecular System

**Figure 1.** Genotypes of RFLP's. "1" indicates presence and "2" indicates absence of the respective restriction site. A: *Taq* I, B: *Hinc* II, C: *Ava* II.

microsatellite marker tandem repeat  
number (Figure 2). 가 RFLP  
PCR Taq I 6  
5 Hinc II Ava II 37 6  
1.5% agarose gel( , Hinc II 3%  
MetaPhor<sup>®</sup> agarose gel(FMC BioProducts Co., Rock-  
land, Maine, U.S.A.)) ethidium  
bromide UV lamp .

(Figure 1).

4. heterozygosity PIC  
allele heterozygosity  
PIC . PIC Botstein  
14.  
$$PIC = 1 - \sum_{i=1}^n P_i^2 - \sum_{i=1}^{n-1} \sum_{j=i+1}^n 2 P_i^2 P_j^2$$
  
Pi: allele frequency of i allele  
Pj: allele frequency of j allele

5. FH 가  
FH 가

haplotype 가  
(recombination)  
,  
.  
1.  
94  
68 : 26, 42.3 ± 9.5 , total cholesterol 192.9 ±  
33.7 mg/dl, triglyceride 145.4 ± 83.2 mg/dl, HDL-  
cholesterol 42.8 ± 11.0 mg/dl .  
2.  
94 allele  
. RFLP Taq I  
allele 가 0.67, Hinc II  
allele 가 0.85 ,  
Ava II allele  
가 0.84 (Table 4). Microsatellite marker  
. D19S394 6가 , ATn 4가 , D19S221 8가  
allele allele  
tandem repeat number Table 5 .  
3. Heterozygosity PIC  
94 heterozy-

**Table 4. Allele frequency of RFLP markers**

	n	Genotype			Allele frequency	
		1/1	1/2	2/2	1*	2**
<i>Taq</i> I	93	7	46	38	0.33	0.67
<i>Hinc</i> II	94	68	23	2	0.85	0.15
<i>Ava</i> II	91	2	26	66	0.16	0.84

1\* digested by each restriction enzyme

2\*\* not digested by each restriction enzyme

**Table 5. Allele frequency and tandem repeat number of microsatellite markers**

n*	D19S394		ATn		D19S221	
	Frequency	Tandem repeat number	Frequency	Tandem repeat number	Frequency	Tandem repeat number
1	0.09	9	0.49	6	0.04	20
2	0.34	10	0.05	7	0.05	21
3	0.50	11	0.45	8	0.09	22
4	0.05	12	0.01	9	0.14	23
5	0.01	13			0.41	24
6	0.01	14			0.09	26
7					0.14	28
8					0.04	29

n\* The arbitrarily defined allele number

**Table 6. Heterozygosity and PIC value of genetic markers**

Marker	Heterozygosity*	PIC
D19S394	0.64 / 0.59	0.558
<i>Taq</i> I	0.51 / 0.44	0.344
<i>Hinc</i> II	0.25 / 0.26	0.223
<i>Ava</i> II	0.28 / 0.27	0.233
ATn	0.56 / 0.55	0.455
D19S221	0.60 / 0.77	0.475

\* Observed / expected

gosity allele heterozygosity, , PIC Table 6 .

4. heterozygosity PIC

가 haplotype cumulative heterozygosity cumulative PIC heterozygo-  
*Hinc* II *Ava* II

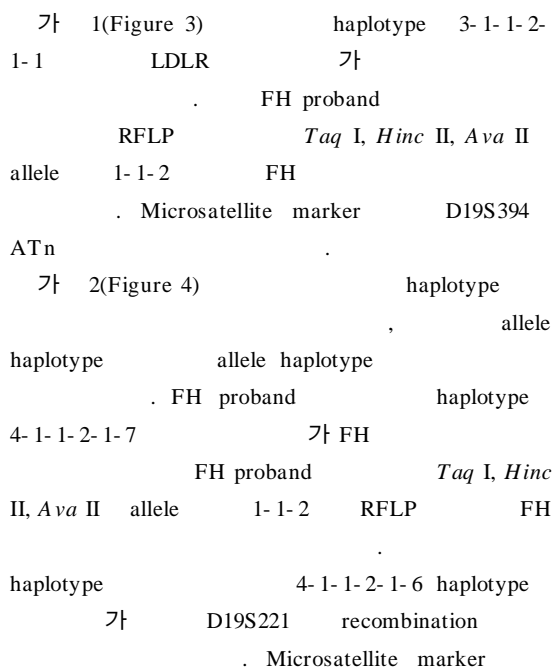
**Table 7. Cumulative heterozygosity and cumulative PIC value of genetic markers**

Marker	Heterozygosity	PIC
<i>Hinc</i> II+ <i>Ava</i> II	0.336	0.331
<i>Taq</i> I+ <i>Hinc</i> II	0.583	0.520
<i>Taq</i> I+ <i>Hinc</i> II+ATn	0.719	0.814
D19S394+ATn	0.813	0.813

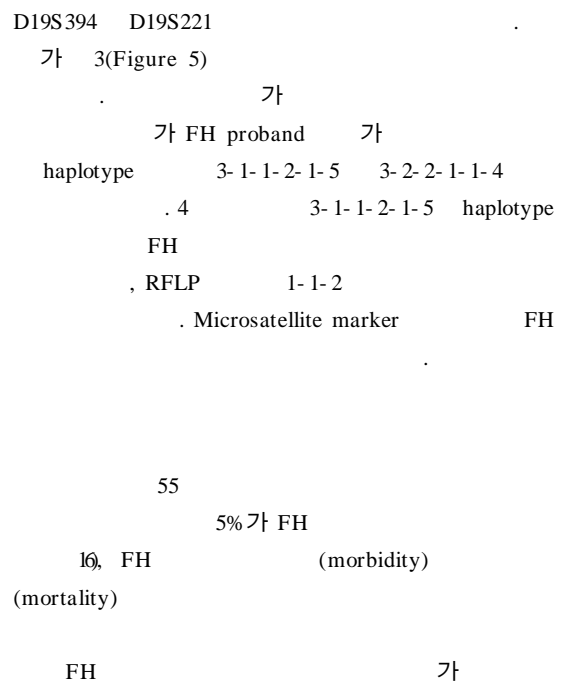
sity 0.336, PIC 0.331 . marker  
(linkage disequilibrium) , |D|  
0.72 ( $p<0.05$ ) 15. *Taq* I *Hinc* II  
heterozygosity PIC 0.583 0.520  
*Taq* I, *Hinc* II, ATn 3가 heter-  
ozygosity PIC 0.719 0.814 .  
D19S394 ATn heterozygosity  
PIC 0.813, 0.813 (Table 7).

5. FH 가 haplotype  
DNA haplotype 가 가 rec-  
ombination 가

**Figure 3.** Haplotype analysis of family 1. Haplotype of the affected gene can be identified as 3-1-1-2-1-1. "1" indicates presence and "2" indicates absence of the respective restriction site in RFLP's. In microsatellite markers the number is arbitrarily defined. The number inside the symbol indicates age.



**Figure 4.** Haplotype analysis of family 2. Haplotype of the affected gene can be identified as 4-1-1-2-1-7. "1" indicates presence and "2" indicates absence of the respective restriction site in RFLP's. In microsatellite markers the number is arbitrarily defined. The number inside the symbol indicates age. \* Recombination is suspected. \*\* Haplotypes are supposed by her offspring's haplotypes.



, 가 RFLP

Weiss	<i>Taq</i> I, <i>Hinc</i> II, <i>Nco</i> I	RFLP
PIC	0.236, 0.367, 0.362	
	0.223- 0.344	RFLP
FH	LDLR	
가	12	
FH 가	RFLP	haplotype
, 가	1 가 3	RFLP 3가
	FH	LDLR
	가	, RFLP
FH		. RFLP
	가	RFLP
	microsatellite marker	
LDLR	microsatellite marker	
D19S394, ATn, D19S221		12 13 18
D19S394 LDLR	telomere	250kb
	Joanne	FH
가	G528D, V408M, S265R,	
C152R LDLR	91- 100% linkage	
	13, ATn LDLR	exon 18
3'		LDLR
tandem repeat	가	

**Figure 5.** Haplotype analysis of family 3. Haplotype of the affected gene can be identified as 3-1-1-2-1-5. "1" indicates presence and "2" indicates absence of the respective restriction site in RFLP's. In microsatellite markers the number is arbitrarily defined. The number inside the symbols indicates age. \* He died from acute myocardial infarction and supposed to be a FH proband. \*\* She is a type II diabetic patient.

	FH	LDLR		recombination microsatellite marker	D19S221	LDLR
		LDLR	300	1.3 Mb	centromere	,
		, 가		가	LDLR	57- 71% linkage dise-
	LDLR		.	equilibrium		I3 I8.
			.	microsatellite marker		PIC
xanthoma		FH	, tendon	D19S394, ATn,	D19S221	0.558, 0.455,
			,	0.475	RFLP	
					0.9, 0.530, 0.8	
Torres		FH	.	I2 I3 I8. Tandem repeat	polymorphism	DNA
		heterozygote	LDLR		replication slippage	
			LDL-		가	,
cholesterol			I7), FH			
		FH		I9)		
			.	FH 가	microsatellite marker	
	FH proband		LDLR		가	가
	FH					, 가



- 7 : 가 -

1 D19S394 ATn 가 2  
D19S394 D19S221  
microsatellite marker : 가 (Familial hypercho-  
PIC 0.5 , lesterolemia, FH) low density lipoprotein  
microsatellite marker receptor(LDLR)  
2가 가  
D19S394 allele 가  
6가 D19S221 8가 가 가 .  
PIC 0.558 3가 microsatellite marker 가  
LDLR .  
가 LDLR : 가 94  
90% linkage disequilibrium LDLR 3가 restriction frag-  
FH Taq I, Hinc  
D19S221 PIC 0.475 II, Ava II, microsatellite marker D19S394, ATn,  
FH 가 haplotype D19S221 allele , heterozygosity, polymor-  
1 recombination phism information content(PIC)  
가 , FH 가  
D19S221 LDLR 1.3 Mb 15 haplotype  
recombination .  
가 FH : 94  
ATn microsatellite marker heterozygosity PIC D19S394, Taq I,  
allele 가 4가 PIC 0.455 Hinc II, Ava II, ATn, D19S221 0.64/0.558,  
LDLR tandem 0.51/0.344, 0.25/0.223, 0.28/0.233, 0.56/0.455, 0.60/0.475  
repeat sequence 2). Hinc II Ava II |D|=0.72( $p<0.05$ )  
가 D19S394 ATn PIC  
PIC 0.813 Taq I, Hinc Taq I+Hinc II가 0.520, Taq I+Hinc II+ATn  
II, ATn PIC 0.814 가 0.814, D19S394+ATn 0.813 . FH가  
haplotype  
RFLP  
3가  
가 , microsatellite marker  
D19S394 ATn 가 가  
D19S221 1 recombination  
: LDLR  
Taq I, FH  
LDLR 가 , 가  
FH 가  
가 , D19S394 ATn  
가 가 .  
가 가 .

## REFERENCES

- 1) Goldstein JL, Brown MS. The LDL receptor defect in familial hypercholesterolemia. *Med Clin North Am* 66:335, 1982
- 2) Horsthemke B, Dunning A, Humpries S. Identification of deletion in the human low density lipoprotein receptor gene. *J Med Genet* 24:144, 1987
- 3) Leitersdorf E, Hobbs HH. Human LDL receptor gene: Hinc II polymorphism detected by gene amplification. *Nucleic Acids Res* 16:7215, 1988
- 4) Yamamoto T, Davis CG, Brown MS, Schneider WJ, Casey ML, Goldstein LL, Russel DW. The human LDL receptor: a cystein rich protein with multiple Alu sequences in its mRNA. *Cell* 39:27, 1984
- 5) Hobbs HH, Brown MS, Goldstein JL. Molecular genetics of the LDL receptor gene in familial hypercholesterolemia. *Hum Mutat* 1:445, 1992
- 6) Day INM, Wilson DJ, Whittall RA, Heath KE, Haddad L, Humpries SE. A mutation databasing model using full sequence display at the core element of presentation: the LDLR gene in familial hypercholesterolemia. Presented at Mutation Database Meetings American Society of Human Genetics, October 29, 1996
- 7) , , , , , , , 가  
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41:274, 1997
- 8) Leitersdorf E, Chakravarti A, Hobbs HH. polymorphic DNA haplotypes at the LDL receptor locus. *Am J Hum Genet* 44:409, 1989
- 9) Daga A, Mattioni T, Balestreri R, Coviello DA, Corte G, Bertolini S. Use of three DNA polymorphism of the LDL receptor gene in the diagnosis of familial hypercholesterolemia. *Hum Genet* 84:412, 1990
- 10) Weber JL, May PE. Abundant class of human DNA polymorphism which can be typed using the polymerase chain reaction. *Am J Hum Genet* 44:388, 1989
- 11) Gyapay G, Morissette J, Vignal A, Dib C, Fizames C, Millaseau P, Marc S, Bernardi G, Lathrop M, Weiss N, Gudnasson, Ostwald P, Humphries S, Schuster H, Keller C. The usefulness of three biallelic restriction fragment length polymorphism versus a polymorphic dinucleotide tandem repeat polymorphism at the low density lipoprotein receptor gene locus for diagnosis of familial hypercholesterolemia. *Dis Markers* 13:141, 1997
- 12) Joanne TS, Nicholas M, Emmanuel K, Eurydiki D, Steve E, Ian NM. Analysis of low density lipoprotein receptor gene mutations and microsatellite haplotypes in Greek FH heterozygous children: six independent ancestors account for 60% of probands. *Hum Genet* 102:343, 1998
- 13) Botstein D, White RL, Skolnick M, Davis R. Construction of genetic linkage map in man using restriction fragment length polymorphism. *Am J Hum Genet* 32:314, 1980
- 14) Philip WH. Gametic disequilibrium measures: Proceed with caution. *Genet* 117:331, 1987
- 15) Herbert S, Friedrich CL. Clinical criteria versus DNA diagnosis in hetrozygous familial hypercholesterolemia. *Arterioscler Thromb Vasc Biol* 18:331, 1998
- 16) Torres AL, Moorjani S, Vohl MC, Gagne C, Lamarche B, Brun LD, Lupien PJ, Despres JP. Heterozygous familial hypercholesterolemia in children: low density lipoprotein receptor mutation analysis and variation in the expression of plasma lipoprotein-lipid concentrations. *Atherosclerosis* 126:163, 1996
- 17) De Oliveira Silva ER, Haddad L, Kwiterovich PO Jr, Humphries SE, Day INM. Applicability of LDLR flanking microsatellite polymorphism for prenatal diagnosis of homozygous state for familial hypercholesterolemia. *Clin Genet* 53:375, 1998
- 18) Schlottterer C, Trautz D. Slippage synthesis of simple sequence DNA. *Nucleic Acids Res* 20:211, 1992
- 19) Zaliani G, Hobbs HH. Dinucleotide repeat polymorphism at the 3' end of the LDLR gene. *Nucleic Acids Res* 18:4300, 1990